

SHORT COMMUNICATION

EVIDENCE FOR D-CONFIGURATION OF N-MALONYLMETHIONINE, THE METABOLITE OF D-METHIONINE IN *NICOTIANA RUSTICA**

B. LADEŠIĆ, M. POKORNY and D. KEGLEVIĆ

Tracer Laboratory, Institute "Ruder Bošković", Zagreb, Yugoslavia

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Abstract—Evidence for the D-configuration of *N*-malonylmethionine-methyl-¹⁴C isolated from tobacco plant fed with D-methionine-methyl-¹⁴C was obtained by acid hydrolysis of the conjugate and subsequent deamination of methionine with D-amino acid oxidase. Parallel experiments with synthetically prepared *N*-malonyl-D-methionine were carried out.

OUR STUDIES on the metabolism of optical isomers of methionine in tobacco plant have shown¹ that, under identical experimental conditions, the D-isomer is metabolized into the corresponding *N*-malonyl conjugate while this pathway is lacking with its enantiomorph. In the present communication, evidence for the D-configuration of *N*-malonylmethionine formed in tobacco plant is presented.

A large amount of D-methionine-methyl-¹⁴C was applied to five intact *Nicotiana rustica* plants and *N*-malonylmethionine-methyl-¹⁴C was isolated from the root extracts by the procedure described previously.¹ Chromatographically and radiochemically pure metabolite was subjected to acid hydrolysis and aliquots were tested as substrates for L- and D-amino acid oxidase. For comparison, *N*-malonyl-D-methionine was prepared and all assays on the metabolite were carried out in parallel with the synthetically obtained conjugate. Since, α -keto- γ -methylthiobutyric acid, the deamination product of methionine, is an unstable compound especially susceptible to oxidation to the sulfoxide stage¹⁻³ the amount of methionine in the incubation mixture before and after the enzymatic reaction was quantitatively determined.

The results of the experiments clearly show that methionine originating from the *N*-malonyl conjugate formed in the plant is not a substrate for L-amino acid oxidase while with D-amino acid oxidase it undergoes a complete deamination (Table 1). The resulting keto acid was identified as the 2,4-dinitrophenylhydrazone derivative. However, during the isolation procedure, some decomposition of the acid took place and a great part of the hydrazone,

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¹ D. KEGLEVIĆ, B. LADEŠIĆ and M. POKORNY, *Arch. Biochem. Biophys.* **124**, 443 (1968).

² T. A. LARUE, B. F. GERULAT and C. P. BERG, *Arch. Biochem. Biophys.* **121**, 22 (1967).

³ M. POKORNY, E. MARČENKO and D. KEGLEVIĆ (in press).

when chromatographed on paper, underwent oxidation to the corresponding sulfoxide; the radioactivity of both the hydrazones accounted for 61–67% of the initial substrate activity.

The present finding is an additional support for the presumption^{1,3,4-6} that *N*-malonylation of all D-amino acids in plants proceeds under retention of configuration.

TABLE 1. OXIDATION OF METHIONINE BY THE ACTION OF D-AMINO ACID OXIDASE

Sample originating from	Amount tested		Amount recovered		Total deamination calculated	
	I (μg)	II (counts/min)	I (μg)	II (counts/min)	I (%)	II (%)
<i>N</i> -Malonylmethionine-methyl- ¹⁴ C, from plant	232	14,100	8	630	96.6	96.2
	224	13,620	10	650	95.5	95.2
<i>N</i> -Malonyl-D-methionine, synthesized	270	—	10	—	96.2	—
	270	—	13	—	95.2	—
D-Methionine	150	—	4	—	97.3	—

EXPERIMENTAL

Materials and Methods

D-Methionine-methyl-¹⁴C (90 mg, spec. act. 56.14 μC/m-mole, [α]_D +9.9°, in water) was administered in one dose to five hydrocultured intact *Nicotiana rustica* plants which were then allowed to metabolize for 6 days. *N*-Malonylmethionine-methyl-¹⁴C was isolated from the MeOH root extracts as already described,¹ the final purification of the metabolite being performed¹ by fractionation on a Sephadex G-25 column. D-Amino acid oxidase (from hog kidney, Fluka, pract. and Koch-Light 16 units/mg) and L-amino acid oxidase (from *Crotalus adamanteus*, Koch-Light) were used.

Paper chromatography, electrophoresis and radioactive counting procedures were performed as already described.¹ Methionine was determined by the ninhydrin method following essentially the standard procedure;⁷ extinction was measured on a Beckman DU spectrophotometer at 510 mμ.

Configuration of C-2 in *N*-Malonylmethionine

Samples of the conjugate were hydrolysed in 2 N HCl at 100° for 2 hr. Hydrolysates were evaporated to dryness *in vacuo*, the residues were dried over NaOH and dissolved in water to a known volume. Portions of each sample were applied to Whatman paper, previously washed with *n*-BuOH-acetic acid-H₂O (60:15:25), chromatographed in the same solvent and scanned for radioactivity; the methionine peak was either eluted with 50% EtOH and checked for radioactivity, or developed with ninhydrin (0.5% in acetone-0.05 M phosphate buffer pH 7.0, 9.5:0.5, v/v), the spot eluted with EtOH-H₂O (3.5:1, v/v plus 5 mg CuSO₄ × 5H₂O) and the extinction of the solution was measured.

Incubations were performed with 1–2 μmole of methionine, 0.05 M pyrophosphate buffer, pH 8.2, and 0.7–1.0 mg of D-amino acid oxidase in a total volume of 2.0 ml at 37° for 10–16 hr. Reaction mixtures were deproteinized by addition of 10 ml EtOH, followed by centrifugation. Aliquots of the supernatant were evaporated to dryness *in vacuo*, the residue was dissolved in about 1 ml of water and passed through a column of Dowex 50X-8 H⁺ followed by water. Elution was carried out with N NaOH, the amino acid fraction was neutralized with 0.1 N acetic acid and concentrated *in vacuo* to known volume: aliquots were then taken for spectrophotometric and radioactive determination as described above. Aliquots of the supernatant or water effluent were shaken with 0.2% 2,4-dinitrophenylhydrazine solution in 2 N HCl, the formed hydrazones were isolated by extraction procedure¹ and checked for radioactivity and chromatographic purity.

The same assays with synthetically prepared *N*-malonyl-D-methionine, as well as control experiments in which methionine and D-amino acid oxidase were omitted together and separately, were performed in parallel.

Syntheses

N-Ethoxymalonyl-D-methionine methyl ester was prepared from D-methionine methyl ester⁸ and monoethyl malonate with dicyclohexylcarbodi-imide as already described for the DL-compound.¹ Viscous oil, yield

⁴ N. ROSA and A. C. NEISH, *Can. J. Biochem.* **46**, 797 (1968).

⁵ W. ESCHRICH and T. HARTMANN, *Planta* **85**, 213 (1969).

⁶ B. LADEŠIĆ and D. KEGLEVIĆ, *Phytochem.* **8**, 51 (1969).

⁷ V. M. INGRAM and A. O. W. STRETTON, *Biochim. Biophys. Acta* **63**, 20 (1962).

⁸ M. BRENNER, H. R. MÜLLER and R. W. PFISTER, *Helv. Chim. Acta* **33**, 568 (1950).

90%, $[\alpha]_D +7.8^\circ$ (ca. 2.0 in ethanol); for analysis it was distilled in a micro-tube at $145-150^\circ/0.01$ mm Hg (Al block). (Found: C, 47.41; H, 6.92; N, 5.25; S, 11.49. $C_{11}H_{19}NO_3S$ required: C, 47.64; H, 6.90; N, 5.05; S, 11.56%.)

N-Malonyl-D-methionine was obtained as described for the DL-compound;¹ viscous oil (yield 90%) which solidified on standing; $[\alpha]_D -17.2^\circ$ (ca. 2.0 in ethanol). *Bis*-dicyclohexylamine salt: m.p. $142-143^\circ$, from ethanol-ether, $[\alpha]_D -17.5^\circ$ (ca. 1.6 in ethanol). (Found: C, 64.41; H, 10.06; N, 7.20; S, 5.36. $C_{32}H_{59}N_3O_3S$ required: C, 64.28; H, 9.95; N, 7.03; S, 5.36%.) Ca-salt: white powder containing 1 mol. H_2O $[\alpha]_D -10.0^\circ$ (ca. 1.7 in water). (Found: C, 32.78; H, 4.52; N, 4.80; Ca, 13.38. $C_8H_{11}NO_3SCa \times H_2O$ required: C, 32.98; H, 4.49; N, 4.81; Ca, 13.76%.)

N-Malonyl-D-methionine sulfoxide, Ca-salt. A solution of *N*-malonyl-D-methionine Ca-salt (136 mg) in 1 ml of water was oxidized with 0.05 ml of 30% H_2O_2 at room temp. for 3 hr. After evaporation the residue was crystallized from 60% EtOH; on standing at 0° , 109 mg (73%) of white crystals with $[\alpha]_D -20.0^\circ$ (ca. 1.6 in water) separated. The compound crystallized with 1.5 H_2O . (Found: C, 30.45; H, 4.51; N, 4.56; Ca, 12.98. $C_8H_{11}NO_3SCa \times 1.5 H_2O$ required: C, 30.37; H, 4.46; N, 4.43; Ca, 12.66%.)

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